# Accutest® URS-2 Urine Reagent Strips

## **Intended Use**

Accutest®URS-2 Urine Reagent Strips for Urinalysis are in vitro diagnostic test devices that use reagents for qualitative and semi-quantitative urinalysis. Accutest® URS-2 Urine Reagent Strips are for single use in professional near patient (point-of-care) facilities and centralized laboratory locations by medical technologists read visually.

Accutest® URS-2 Urine Reagent Strips for Urinalysis are intended for use to detect conditions indicating possible diabetes, metabolic abnormalities, liver diseases, kidney function, and urinary tract infections. Test results can be used along with other diagnostic information to rule out certain disease states and to determine if microscopic analysis is needed.

### **Summary and Explanation of Tests**

Accutest® URS-2 Urine Reagent Strips provide tests for Glucose and Protein in Urine.

### **Test Principles**

**Protein**: The test is based on the protein-error-of-indicators principle. An ion in the specific pH indicator attracted by a cation on the protein molecule makes the indicator further ionized, which changes its color.

**Glucose**: The glucose oxidized by glucose oxidase catalyzes the formation of glucuronic acid and peroxide hydrogen. Peroxide hydrogen releases neo-ecotypes oxide [O] under the function of peroxidase. [O] oxidizes iodide potassium, which causes the color change.

## Reactive Ingredients (based on dry weight at time of impregnation)

**Protein**: 0.1% w/w tetrabrompenol blue; 97.4% w/w buffer; 2.5% w/w nonreactive ingredients.

**Glucose**: 1.7% w/w glucose oxidase (microbial, 123U); 0.2% w/w peroxidase (horseradish, 203U); 71.8% w/w buffer; 0.1% w/w potassium iodide; 26.2% w/w nonreactive ingredients.

## Storage

Strips must be kept in the original bottle. Transfer to any other container may shorten the expiration date of product. Store at temperatures between 2-30 degrees C (39-86 degrees F). Keep away from direct sunlight and moisture. Do not remove desiccants in the bottles. Replace the cap immediately after removing reagent strips. Protect against exposure to light, heat, and ambient moisture to guard against altered reagent reactivity.

## **Specimen Collection and Preparation**

Collect fresh urine in a clean and dry container. Do not centrifuge the urine. Mix the sample well before testing it [1]. The container should allow for complete dipping of all reagent strip areas. Test the urine within four hours after voiding, sooner if testing for bilirubin or urobilinogen [2].

#### **Expected Results**

The sensitivity of Accutest®URS-2 Urine Reagent Strips for Urinalysis in testing clinical urine specimens may vary depending upon several factors, such as the variability of color perception, specific gravity, pH values, and the lighting conditions when strips are read visually. Visual reading results may not exactly match instrumental reading results because of the difference between the perception of human eyes and the optical instrument. Most visual and instrument readings are within one level of the true value.

#### **Procedure**

## **Gather Materials**

- Dry and clean plastic container
- > Toilet paper
- Watch with a second hand or stopwatch (if you read the strip visually)
- Urinalysis reagent strips

#### **Perform Tests**

- Immerse the reagent area of the strip in the urine specimen and take it up quickly and immediately. Start timing if reading visually.
- 2. Run the edge of the strip against the rim of the container to remove excess urine. Lay the strips on a paper towel with the reagent areas upward.
- 3. Hold the strip up horizontally and compare the reagent areas on the strip to the corresponding color chart on the bottle label at the exact times specified and in good light. Hold the strips close to the color blocks and match carefully. Make note of the result. Color changes after 2 minutes are of no diagnostic value. If reading by instrument, carefully follow the directions given in the instrument operating instruction. The instrument will automatically read each reagent area at a specified time.
- 4. Dispose of strips with laboratory waste. Do not flush down toilet.







## **Quality Control**

Remove one strip from the bottle and check against the color blocks on the color chart. If the color of the reagent area is darker than the lowest block on the chart (except for specific gravity and pH), the strip is unusable. Discard the strip and check all strips from the bottle before using or discard the bottle. When a new bottle is first opened, use two strips to test known negative and positive specimens or controls. Water should NOT be used as a negative control.

#### **Important Notes**

- Do not take the strips from the bottle unless they are for immediate use.
- Do not touch reagent areas of strips.
- 3. Do not use strips beyond the expiration date.
- Each strip can be used only once.

- Large amounts of ascorbic acid may effect the test for glucose. [2,4].
- Deterioration may result in discoloration or darkening of the reagent areas of the strip. If this happens, or the test results are questionable or inconsistent with expected results, check and make sure the strips are within the expiration date, and also check results with the control urine.

#### **Limitations**

**Protein**: False positive results may be obtained with highly buffered or alkaline urines. Contamination of the urine specimen with quaternary ammonium compounds (e.g., from some antiseptics and detergents) or with cleansers containing chlorhexidine may also produce false positive results [2,4].

**Glucose**: Ascorbic acid concentrations of 10.2 mg/dL and/or acetoacetic acid concentrations of 19.4mg/dL or lower will not influence the test [2].

## **Expected Values/Reference Ranges**

Expected values for a "normal" healthy population and abnormal populations are listed below for each test. Expected values are referenced to European Urinalysis Guidelines, The Clinical Analysis Of Urine Recent Period and Compendium – Urinalysis With Test Strips [2,4,5].

**Protein**: The reagent area is more sensitive to albumin than to globulins, hemoglobin, Bence-Jones protein, and muco-protein. Therefore a 'Negative Result' is not sufficient to indicate that these proteins do not exist in urine. Normally protein is not detectable in urine with conventional methods, although a minute amount of protein is excreted through normal kidney function. Protein in urine is indicated when the color is darker than the plus/minus mark on the chart.

**Glucose**: Normally, a small amount of glucose may be excreted through the kidneys. The amount is usually below the sensitivity of the reagent test. Results at the first positive level may be significantly abnormal if found consistently.

### **Performance Characteristics**

The performance characteristics of the strips are determined by clinical analysis and study. The results from visual readings and represent an actual range of analyte concentrations. Because of the variety of the specimens and reading methods, the values obtained from the results of tests may have errors compared to the actual values of the specimens. Visual reading results may not exactly match the instrumental reading results because of the inherent difference between the perception of human eyes and the optical instruments.

The following table shows the +/-1 color block % Agreement using 1514 samples in laboratory comparison studies between Accutest® URS-2 Urine Reagent Strips and Bayer Multistix 2 SG Reagent Strips.

<u>Analyte</u> % Agreement Glucose 96.9 % (1467/1514) Protein 99.9 % (1513/1514)

**Protein**: In 90% of urines tested, albumin concentrations of 0.15 g/L or greater will produce a color change. The test pad is more sensitive to albumin than globulin, Bence-Jones proteins, and mucoproteins.

**Glucose**: In 90% of urines tested, glucose concentrations of 80 mg/dL or greater will produce a positive result. Sugars other than glucose will not react with the reagent. If the color appears somewhat mottled at the higher glucose concentrations, match the darkest color to the blocks.

The sensitivity of the strips on clinical urine specimens may vary depending upon several factors, such as the variability of color perception, specific gravity, pH value, and the lighting conditions when the strips are read visually. Test sensitivities and output values are given in the following table.

### Sensitivity and Output Values of Accutest®URS-2 Urine Reagent Strips

Test Pad	Sensitivity	Output Value
		Visual Read
Protein (mg/dL)	15-30	Neg - 2000
Glucose (mg/dL)	50-100	Negative - 2000

## **Bibliography**

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- 2. "European Urinalysis Guidelines", The Scandinavian Journal of Clinical & Laboratory Investigation, Scand J Clin Lab Invest-Vol. 60-Supplement 231.2000.
- 3. "Operating Rules Of Clinical Test" (Rev.2), The Ministry of Health of P.R.C. Publishing. Yingwu Ye, Yusan Wang.
- "The Clinical Analysis of Urine Recent Period", The Science and Technology Publishing House, Yu Long Cong, Jun Long Ma, Editors; 1998; pp. 37-81, 96-97.
- 5. "Compendium Urinalysis with Test Strips" Roche Diagnostic, Combur® Reagent Strips.



# Notes on Symbols and Marks



16530 Ventura Blvd. #512, Encino, CA 91436 Toll Free: 800-676-5565 Phone: 818-986-8530 Fax: 818-986-0235 Rev.04/2013